

Biology 5357
Chemistry & Physics of Biomolecules
Examination #1

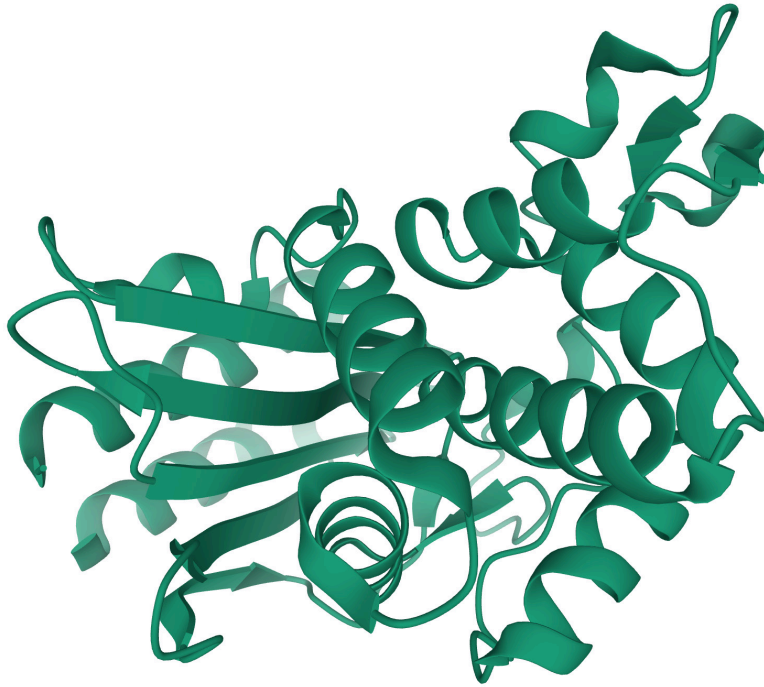
Proteins Module

September 30, 2022

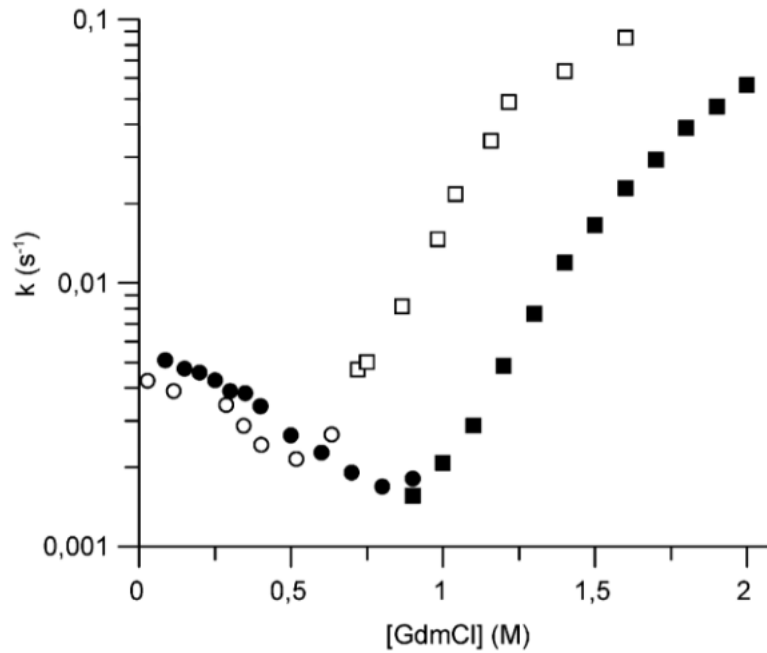
Name: _____

Question 1 (30 points; A-C = 10 pts each)

- (A) Analyze the structure shown below of the TEM-1 β -lactamase protein, which is derived from the PDB entry 5HVI. For example, what kinds of secondary structural elements and what motifs does it contain. Comment on packing, twist, *etc.* of the various elements. What top-level class of protein fold does the structure belong to?



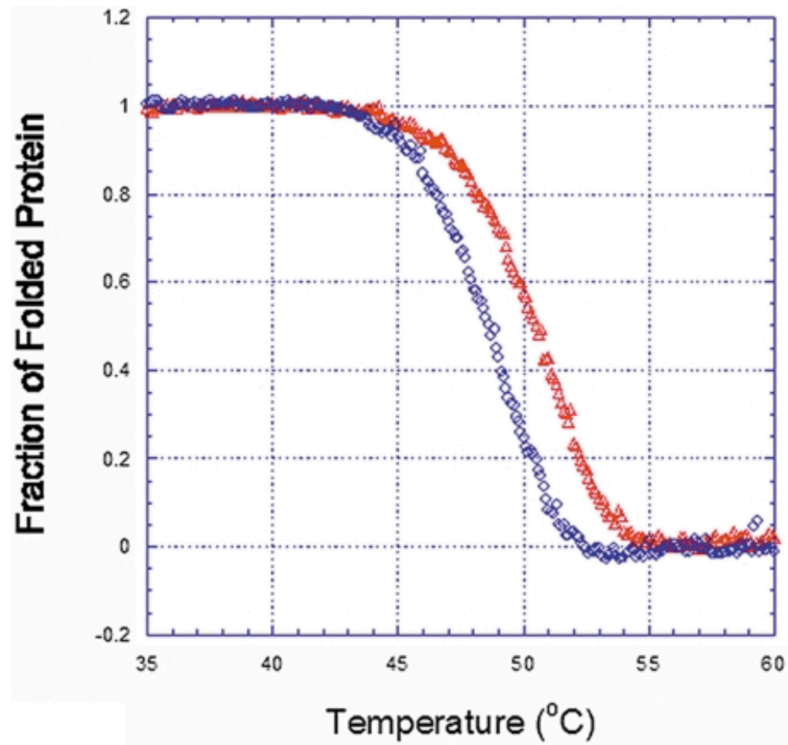
(B) The plot below shows data from kinetic folding-unfolding experiments on wild-type (filled symbols) and the W290F mutant (empty symbols) of β -lactamase. Explain the difference between points represented by circles and those represented by squares. What do the plots indicate about relative rates of folding and unfolding for the wild-type and mutant? What does the data suggest about involvement of residue 290 in the folding path of β -lactamase?



- (C) “Chevron plots” demonstrate that rates of folding (k_f) and unfolding (k_u) vary with denaturant concentration, [d]. Let m_f and m_u represent the slope of the folding and unfolding arms of a typical 2-state chevron plot. Suggest an equation that gives the equilibrium folding-unfolding free energy (ΔG) as a function of denaturant concentration [d] and the two slopes, m_f and m_u .

Question 2 (20 points; A-B = 10 pts each)

The plot below shows equilibrium thermal unfolding data for TEM-1 β -lactamase. The figure is taken from a paper by Horn and Shoichet (*Journal of Molecular Biology*, **336**, 1283-1291, 2003). The red curve corresponds to the *apo* β -lactamase protein, while the blue curve is with a non-competitive inhibitor bound.



- (A) Analyze the data to estimate the ΔG value between folded and unfolded forms for both the *apo* and ligated protein, assuming for the moment that ΔC_p is zero. Additionally, find the ΔS associated with the folding-unfolding process.

- (B) The heat capacity change, ΔC_P , associated with protein unfolding is not zero, but can be estimated from an approximate formula provided by Pace and Scholtz in their review *Measuring the Conformational Stability of a Protein*:

$$\Delta C_P = 172 + (17.6 \times N) - (164 \times SS)$$

In this equation, ΔC_P is the heat capacity change in cal/mol/K, N is the number of residues in the protein, and SS is the number of disulfide bonds in the structure. Note that β -lactamase has 263 residues, and contains a single disulfide bond between residues 77 and 123. What is the estimated ΔC_P value for β -lactamase? Why might the equation contain a term involving the number of disulfide bonds?

The ΔC_P value can then be used to compute ΔG at different temperatures, T , using another formula given in the Pace article:

$$\Delta G(T) = \Delta H_m (1 - T/T_m) - \Delta C_P [(T_m - T) + T \ln(T/T_m)]$$

How could you use this approach to find the stability of *apo* β -lactamase at room temperature (25°C) and at physiological temperature (37°C)? *You do not need to actually perform the calculations. Simply describe in words the steps you would follow to estimate the stabilities.*

Question 3 (12 points; A-E=4 pts each)

Briefly discuss and explain the following statements of about protein folding.

- (A) “The potential energy surface for a protein is funnel-shaped.”

- (B) “The hydrophobic effect is a driving force leading to globular structures.”

- (C) “Protein folding is a highly cooperative process.”

Question 4 (8 points; A & B=3 pts each, C=2 pts)

- (A) A typical drug inhibiting a target protein might have a dissociation constant, K_D , of 2.5 nM. Write the equation for the equilibrium represented by K_D .

- (B) What is the binding free energy, ΔG , corresponding to $K_D = 2.5$ nM?

- (C) For this value of K_D , at what drug concentration would the protein be half-saturated by drug?

Question 5 (10 points; A & B=3 pts each, C & D=2 pts)

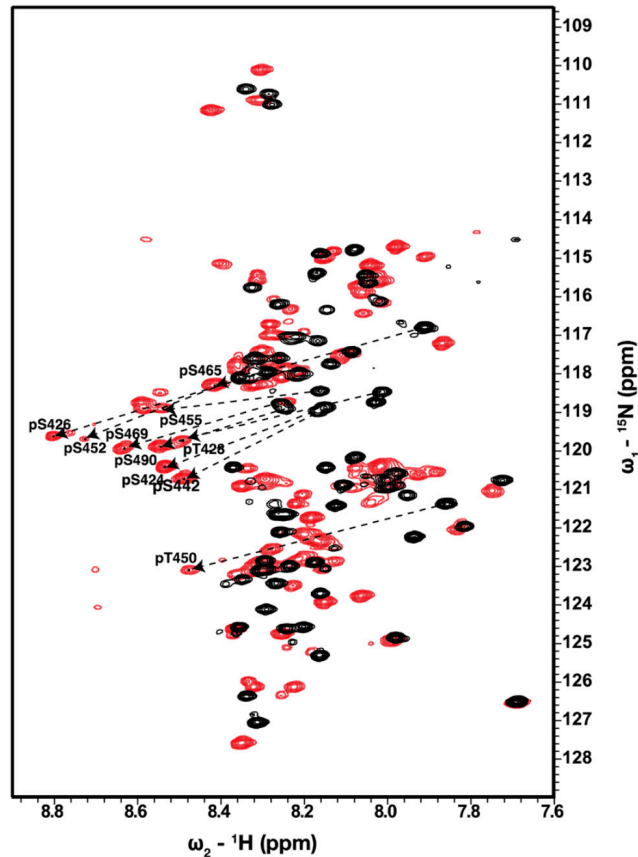
- (A) In general, flexible polymers are well-described by an equation that relates the chain dimension (*e.g.*, radius of gyration R_g) to some other parameters via the equation:

$$R_g = B_0 N^{\nu}$$

What are the meaning of B_0 , N and ν ? [For up to 3 bonus points, indicate the values ν can take in different situations?]

- (B) Greg wants to study the binding of a disordered protein to a partner? Name a technique could Greg use and describe (very broadly) how it works. [You are free to name any technique you feel comfortable discussing.]
- (C) Name one advantage and one disadvantage of single molecule FRET (smFRET) over SAXS.

- (D) The HSQC spectrum below shows a disordered protein prior to (red) and upon (black) phosphorylation. What does each peak represent, and why do some of them move (black dashed lines) upon phosphorylation?



Question 6 (10 points; A=3 & B=3 pts each, C & D=2 pt each)

- (A) Describe three possible functions of disordered regions, explaining how and/or why disorder could be relevant/important in that function?

(B) Disordered regions lack a fixed 3D structure. Despite this, they can engage in specific molecular interactions. In the absence of a fixed 3D structure, how might disordered regions confer specificity?

(C) Disordered regions are always more expanded than folded domains. Is this statement *True* or *False*. Briefly explain.

(D) What would be the best technique to measure local secondary structure in an IDR?

Question 7 (10 points; A=2 pt, B=4 pts, C=4 pts)

- (A) What is a disordered protein or protein region?
- (B) What features of a protein sequence might influence or determine if it is folded or disordered? Name two features and briefly explain why each feature could influence whether the protein is disordered or folded.
- (C) What are some challenges in studying disordered proteins experimentally? Name two challenges and explain the molecular origin for this challenge and what kinds of problems it causes.